



Draft Genome Sequences of Three Isolates of *Coniothyrium glycines*, Causal Agent of Red Leaf Blotch of Soybean

Trenna Blagden,^a Andres Espindola,^{a,b} Kitty Cardwell,^{a,b} Alejandro Ortega-Beltran,^c Ranajit Bandyopadhyay^c

^aDepartment of Entomology & Plant Pathology, Oklahoma State University, Stillwater, Oklahoma, USA

^bNational Institute of Microbial Forensics in Food and Agricultural Biosecurity, Oklahoma State University, Stillwater, Oklahoma, USA

^cInternational Institute of Tropical Agriculture, Ibadan, Nigeria

ABSTRACT *Coniothyrium glycines*, the causal agent of red leaf blotch in soybeans, is considered a high-consequence biological agent. With limited genomic information known, there are no molecular genotyping or detection methods available. We report the draft genome sequences of three *C. glycines* isolates, greatly enhancing our knowledge of this species.

The pycnidial state of a fungal species, *Pyrenochaeta glycines*, was first suspected of causing red leaf blotch of soybeans in 1955 in Ethiopia (1). Later, the sclerotial state of a fungus causing leaf spot disease was described as *Dactuliophora glycines* (2). Following extensive phylogenetic analysis, it is now designated *Coniothyrium glycines* (R. B. Stewart) Verkley & Gruyter (3).

C. glycines is found in soybeans in central and southern Africa (1, 4, 5). Previous records indicate that yield losses can reach up to 50% (6). With the United States annually exporting 52.8 million tons of soybeans, a farm cash value of \$35.2 billion in 2015 (<https://www.ers.usda.gov/topics/crops/soybeans-oil-crops/related-data-statistics/>), the threat posed by *C. glycines* could be significant. It is currently unknown if this fungus can survive in the temperate regions of the United States or what the potential damage from its spread to other cultivated legumes would be (7). The availability of genomic sequences would provide valuable knowledge for detection tool development and facilitate our understanding of genetic diversity within *C. glycines*.

Coniothyrium glycines 13, 17, and 18 are pycnidial isolates originating from soybean leaves collected by Olalekan Ayinde in September 2016 at ECOWAS Church Farm in Jos Plateau, Nigeria. Isolates grown on potato dextrose agar (PDA) plates were sent to Oklahoma State University (Stillwater, OK), where single hyphal tip isolation was performed. They were subcultured on 10% V8 juice agar plates and maintained at 20°C in darkness. The agar plate surface was scraped into 1 ml of sterile reverse-osmosis (RO) water to collect pycniospores for DNA extraction. The spore solution was transferred into a Precellys 24 VK05 lysing kit 2.0-ml tube with 10 2.7-mm glass beads. The Precellys 24 tissue homogenizer (Bertin Instruments, Montigny-le-Bretonneux, France) run was two 25-second cycles at 6,500 rpm with a 5-second pause between cycles. The genomic DNA was extracted using the DNeasy plant and fungal extraction kit (Qiagen, Germantown, MD). Sequencing of *C. glycines* 13, 17, and 18 was performed using the Nextera kit on an Illumina MiSeq platform and a rapid sequencing kit (SQK-RAD004) with a FLO-MIN106.1 SpotON flow cell (R9.4) for the Oxford Nanopore MinION system. Assembly using a hybrid approach with SPAdes 3.13.0 (8) was conducted for *C. glycines* 17 and with Canu 1.8 (9) for *C. glycines* 13 and 18. The Illumina reads were quality controlled with FastQC 0.11.3, and the MinION reads were quality controlled with Albacore 2.3.4. All relevant sequencing and assembly statistics are summarized in Table 1. BUSCO version 3.0.2 using the BUSCO fungi_odb9 database was used to

Citation Blagden T, Espindola A, Cardwell K, Ortega-Beltran A, Bandyopadhyay R. 2019. Draft genome sequences of three isolates of *Coniothyrium glycines*, causal agent of red leaf blotch of soybean. *Microbiol Resour Anounc* 8:e00378-19. <https://doi.org/10.1128/MRA.00378-19>.

Editor Jason E. Stajich, University of California, Riverside

Copyright © 2019 Blagden et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Trenna Blagden, trenna.blagden@okstate.edu.

Received 1 April 2019

Accepted 4 September 2019

Published 3 October 2019

TABLE 1 Sequencing and assembly statistics of *C. glycinis* strain genomes

Isolate	Genome coverage (×)	Total no. of MinION reads used	Illumina avg. read length (bp)	Total no. of Illumina reads used (million)	Genome size (Mbp)	No. of contigs	N_{50} (kbp)	G+C content (%)
13	1,722.105	2,020,000	146.39	11.9	32.75	260	520	52.19
17	1,697.311	518,282	142.63	13.7	36.88	309	269	52.19
18	1,691.057	97,848	146.13	10.8	33.10	262	436	52.19

determine completeness of genome coverage and culture identity confirmed by a BLAST+ 2.8.1 (10) search for a 100% match of the internal transcribed spacer (ITS) region of *C. glycinis*. The BUSCO scores for strains 13, 17, and 18 were all above 96% complete genes, and all isolates had the complete ITS region.

Data availability. This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession numbers [SPUQ00000000](https://accession.gtrdb.org/SPUQ00000000) (strain 18), [SPUR00000000](https://accession.gtrdb.org/SPUR00000000) (strain 17), and [SPUS00000000](https://accession.gtrdb.org/SPUS00000000) (strain 13). The versions described in this paper are versions SPUQ02000000, SPUR01000000, and SPUS02000000. The raw sequence data were deposited under SRA accession numbers [SRR8655434](https://accession.gtrdb.org/SRR8655434) to [SRR8655442](https://accession.gtrdb.org/SRR8655442).

ACKNOWLEDGMENTS

The computing for this project was performed at the OSU High Performance Computing Center at Oklahoma State University, supported in part through National Science Foundation grant OAC-1126330. We thank the OSU Microarray Core Facility, which was supported by grants from the NSF (EOS-0132534) and the NIH (2P20RR016478-04, 1P20RR16478-02, and 5P20RR15564-03). We also thank the Oklahoma Soybean Board for financial support (award number RFP 510230).

T.B. procured funding, carried out experiments, coordinated the Illumina sequencing, performed the MinION sequencing, conducted portions of the data analysis, and took the lead in writing the announcement. A.E. assisted with MinION sequencing, assembled the raw reads, and contributed to the final version of the announcement. K.C. maintained the cultures used in experiments. A.O.-B. provided isolates used in experiments. A.O.-B. and R.B. procured isolates of *Coniothyrium glycinis* from soybean plants in the field on the Jos Plateau of Nigeria.

REFERENCES

- Stewart RB. 1957. An undescribed species of *Pyrenochaeta* on soybean. *Mycologia* 49:115–117. <https://doi.org/10.1080/00275514.1957.12024619>.
- Leakey CLA. 1964. *Dactuliothra*, a new genus of mycelia sterilia from tropical Africa. *Trans Br Mycol Soc* 47:341–350. IN349-IN310. [https://doi.org/10.1016/S0007-1536\(64\)80006-1](https://doi.org/10.1016/S0007-1536(64)80006-1).
- de Gruyter J, Woudenberg JHC, Aveskamp MM, Verkley GJM, Groenewald JZ, Crous PW. 2013. Redisposition of phoma-like anamorphs in *Pleosporales*. *Stud Mycol* 75:1–36. <https://doi.org/10.3114/sim0004>.
- Hartman GL, Datnoff LE, Levy C, Sinclair JB, Cole DL, Javaheri F. 1987. Red leaf blotch of soybeans. *Plant Dis* 71:113–118. <https://doi.org/10.1094/PD-71-0113>.
- Punithalingam E. 1990. *Dactuliochaeta glycinis*. In *IMI descriptions of fungi and bacteria*, no. 102. Sheet 1012. CAB International, Wallingford, United Kingdom.
- Datnoff LE, Naik DM, Sinclair JB. 1987. Effect of red leaf blotch on soybean yields in Zambia. *Plant Dis* 71:132–135. <https://doi.org/10.1094/PD-71-0132>.
- Hartman GL, Haudenschild J, Smith KL, Tooley P, Shelton J, Bulluck R, Engle JS, Magarey RD. 2009. Recovery plan for red leaf blotch of soybean caused by *Phoma glycinicola*. US Department of Agriculture, Washington, DC. <https://www.ars.usda.gov/ARSUserFiles/00000000/opmp/Soybean%20RLB%20April%202011.pdf>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive *k*-mer weighting and repeat separation. *Genome Res* 27:722–736. <https://doi.org/10.1101/gr.215087.116>.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <https://doi.org/10.1186/1471-2105-10-421>.