

1 **First Report of *Lasiodiplodia theobromae* Causing Dieback Symptoms on Plantain (*Musa***
2 **AAB subgroup) in Nigeria**

3 Y. A. Kolombia^{1†}, A. E. Alakonya^{1††}, A. Ortega-Beltran¹, D. Amah¹, J. Agogbua², G. Mahuku³
4 and R. Swennen^{4,5}

5 ¹International Institute of Tropical Agriculture, PMB 5320, Ibadan, Oyo State, Nigeria

6 ²Department of Plant Science and Biotechnology, Faculty of Science, University of Port
7 Harcourt, PMB 5323, Choba, Rivers State, Nigeria.

8 ³International Institute of Tropical Agriculture, Plot 25 Light Industrial Area, Coca Cola Rd,
9 Dar es Salaam, Tanzania

10 ⁴International Institute of Tropical Agriculture, Plot 15B Naguru East Road, Upper Naguru,
11 Box 7878, Kampala, Uganda

12 ⁵Laboratory of Tropical Crop Improvement, Department of Biosystems, KU Leuven, Willem
13 de Croylaan 42, 3001 Leuven, Belgium

14 [†]Current address: International Maize and Wheat Improvement Center (CIMMYT), Carretera
15 México-Veracruz Km. 45, El Batán, Texcoco, C.P. 56237, México

16 Corresponding author: A. E. Alakonya: A.Alakonya@cgiar.org; Y. A. Kolombia:
17 Y.Kolombia@cgiar.org

18 Bananas (banana and plantains) rank sixth among staple food crops (FAO 2018), with
19 production challenged by biotic factors, mainly fungal diseases that may cause a total loss in
20 some orchards (Jones 2018).

21 In April 2017, dieback symptoms (progressive blackening and necrotic aerial plant
22 parts, leaves, fruits and peduncles) were observed on plantain (*Musa* AAB subgroup), in Onne,
23 Rivers State, Nigeria (4°42'55.4012"N, 7°10'35.92128"E). Diseased plants (n=112) were either
24 wilted with blackened necrotic areas, or dead (Fig. S1). Nearly 10% of the plants had blackened
25 pseudostems and fruits with slate gray to black internal tissues when sliced (Fig. S1) and black,
26 erumpent pycnidia were observed on diseased fruits.

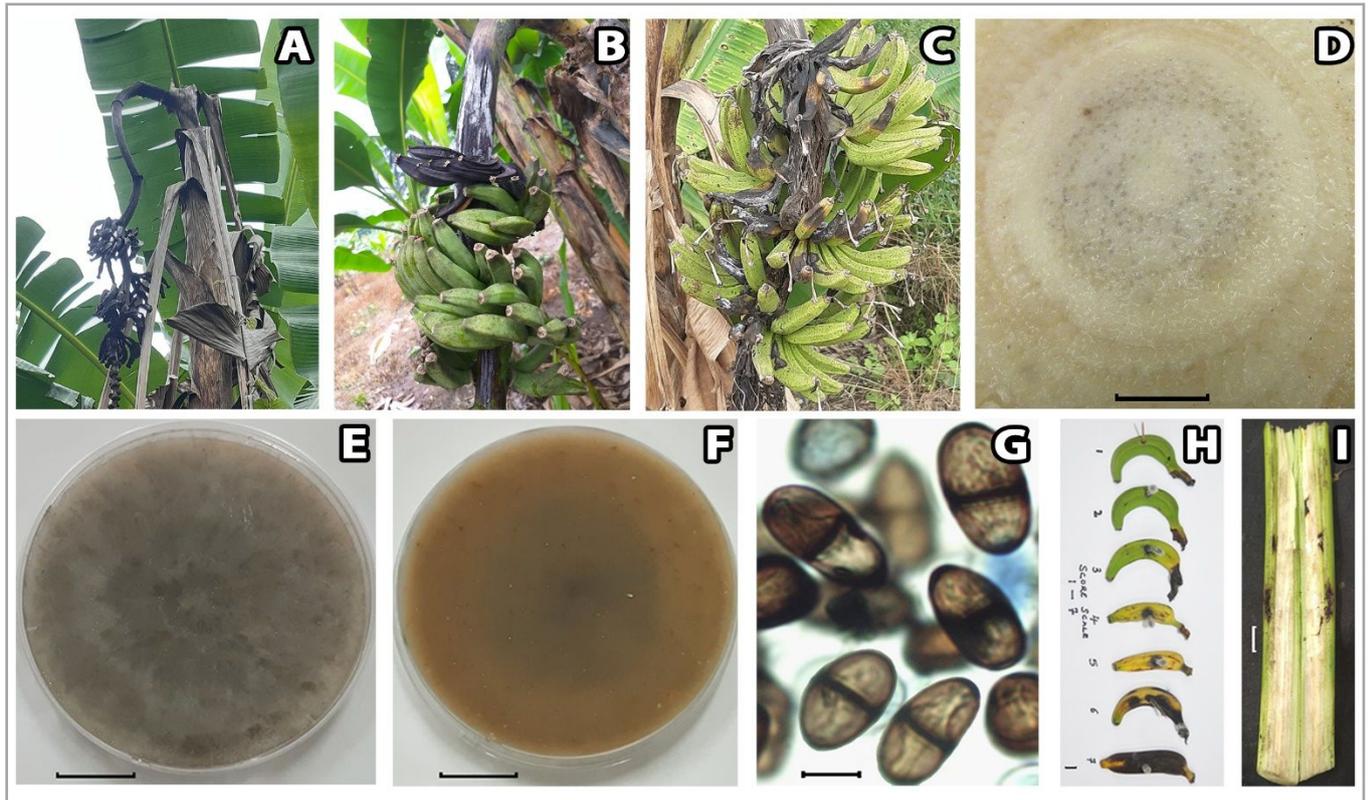
27 A fungal species was consistently isolated when surface disinfected pieces of diseased
28 samples were cultured on PDA plates. Plates were incubated at 25±2°C for 4 to 15 d to observe
29 conidia. Isolates had colonies and conidia consistent with members of the Botryosphaeriaceae
30 family (Phillips et al. 2013). Immature conidia were single-celled, ellipsoidal and hyaline while
31 mature conidia were two-celled, had a thick wall, a central septum, longitudinal striations, and
32 a dark brown, cinnamon-like color. Size of mature conidia (n = 20) ranged 22.9 to 30.0 × 14.2
33 to 18.4 µm ($x = 27.0 \times 15.6$ µm; Fig. S1). DNA templates of three isolates (23688-2_R16;
34 19144-18_R15 and PITA_22-1) were amplified using primers ITS1 and ITS4 for the ITS locus,
35 EF1-688F and EF1-1251R for the translation elongation factor 1- α (TEF-1 α) locus (Phillips et
36 al. 2013) and sequenced (GenBank accession Nos. MZ413346, MZ413347, and MZ413348 for
37 ITS; and MZ420177, MZ420178, and MZ420179 for TEF-1 α). BLASTn query showed 100%
38 identity with reference sequences of various isolates of *Lasiodiplodia theobromae*. Based on
39 morphological characters and nucleotide homology, the isolates were identified as *L.*
40 *theobromae* (Fig. S1 & S2). To fulfil Koch's postulates, 4-month-old plants of plantain hybrid
41 PITA 24, and mature fruits from three genotypes (PITA 24, plantain cultivar Obino L'ewai)
42 were inoculated with mycelial plugs from the margins of 5-d-old cultures of the three *L.*

43 *theobromae* isolates. Pseudostems were drilled with a sterile 5 -mm cork borer, a mycelial plug
44 placed down into the wound, covered with sterilized cotton, and sealed with parafilm. Sterile
45 water was injected every third day to maintain moisture at the inoculated area. Toothpicks
46 containing mycelia were used to inoculate fruits, placed in plastic Crisper boxes. Sterile PDA
47 plugs or toothpicks were used for the controls. Inoculated plants and fruits were kept in a
48 screenhouse at room temperature (~26°C) for 14 d. All inoculated materials developed
49 symptoms similar to the diseased plants in the field. Control plants and fruits remained
50 asymptomatic. *L. theobromae* was re-isolated from the artificially inoculated plant parts and
51 its identity was confirmed. The fungus *L. theobromae* is distributed in tropical and subtropical
52 regions and has a wide host range (Phillips et al. 2013; Mehl et al. 2017). This fungus was
53 previously reported in grey literature as the causal agent of *Musa* spp. basal rot at Onne, Nigeria
54 (Mwangi et al. 2005) but its molecular identification was not conducted; it was unknown
55 whether the isolates were indeed *L. theobromae* or other cryptic species (*L. pseudotheobromae*
56 or *L. parva*) (Alves et al. 2008). Over 15 years later, the present study confirms *L. theobromae*
57 as the causal agent of basal rot of bananas based on nucleotide homology, and to our
58 knowledge, this is the first report of *L. theobromae* causing dieback disease on plantain in
59 Nigeria and in Africa. There is need to conduct a more comprehensive distribution surveys and
60 develop appropriate control strategies in Nigeria.

61 **References**

- 62 Alves, A., Crous, P. W., Correia, A., and Phillips, A. J. L. 2008. Morphological and molecular
63 data reveal cryptic speciation in *Lasiodiplodia theobromae*. *Fungal Divers.* 28:1–13.
- 64 FAO. 2018. FAOSTAT. Food and Agriculture Organization of the United Nations, Rome,
65 Italy. Available at: <http://www.fao.org/faostat/en/> [Accessed August 15, 2020].
- 66 Jones, D. R. 2018. *Handbook of Diseases of Banana, Abacá and Enset.*

- 67 Mehl, J., Wingfield, M. J., Roux, J., and Slippers, B. 2017. Invasive everywhere?
68 Phylogeographic analysis of the globally distributed tree pathogen *Lasiodiplodia theobromae*.
69 Forests. 8:1–22.
- 70 Mwangi, M., Bandyopadhyay, R., Tenkouano, A., and Faturoti, T. 2005. Outbreak of basal end
71 rot on banana and plantain in Nigeria. In *African Crop Science ptoceedings*, p. 293–295.
- 72 Phillips, A. J. L., Alves, A., Abdollahzadeh, J., Slippers, B., Wingfield, M. J., Groenewald, J.
73 Z., et al. 2013. The Botryosphaeriaceae: Genera and species known from culture. *Stud. Mycol.*
74 76:51–167.
- 75

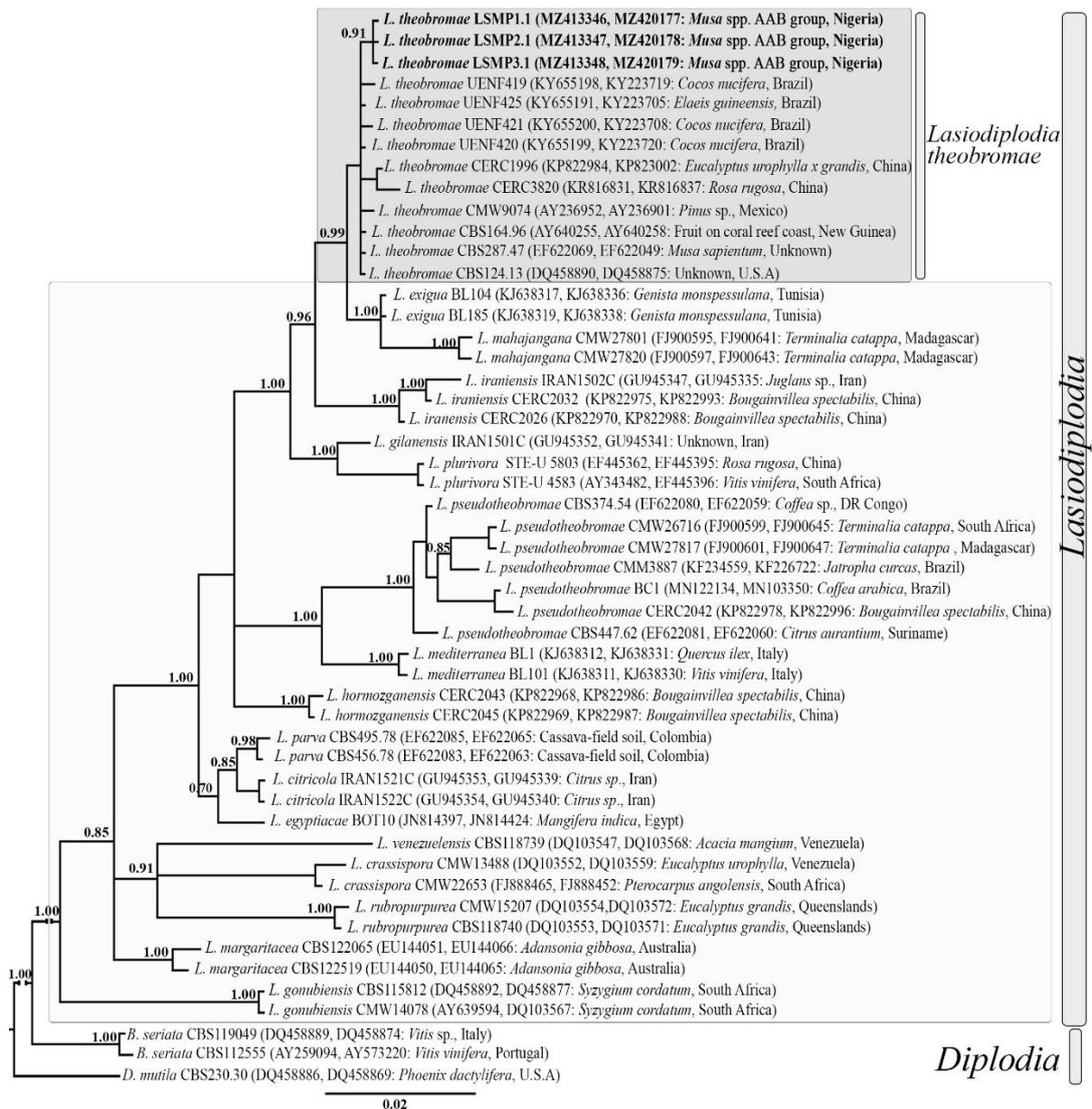


76

77 **Supplementary Figure S1.** Plants with fingers and peduncles rot, dieback symptoms caused78 by *Lasiodiplodia theobromae* (A-C), Pseudostem cross-section of infected plant (D), cultures79 of isolates on PDA (E-F), spores' morphology (G), and the rot caused by *L. theobromae*

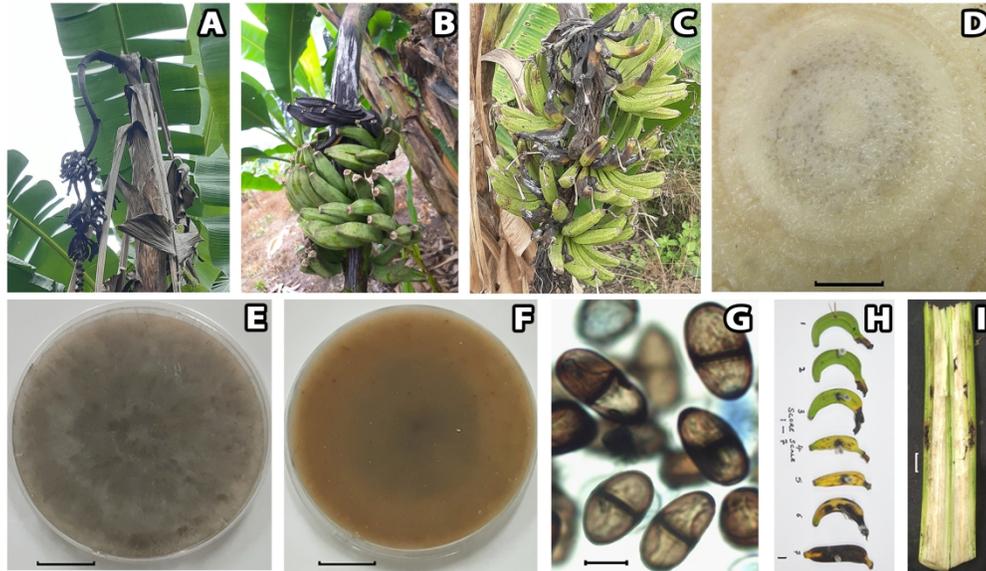
80 following the pathogenicity test on fruits (H) and pseudostems of plants (I). Scale bars: D-F,

81 H-I = 2 cm; G = 10 μ m



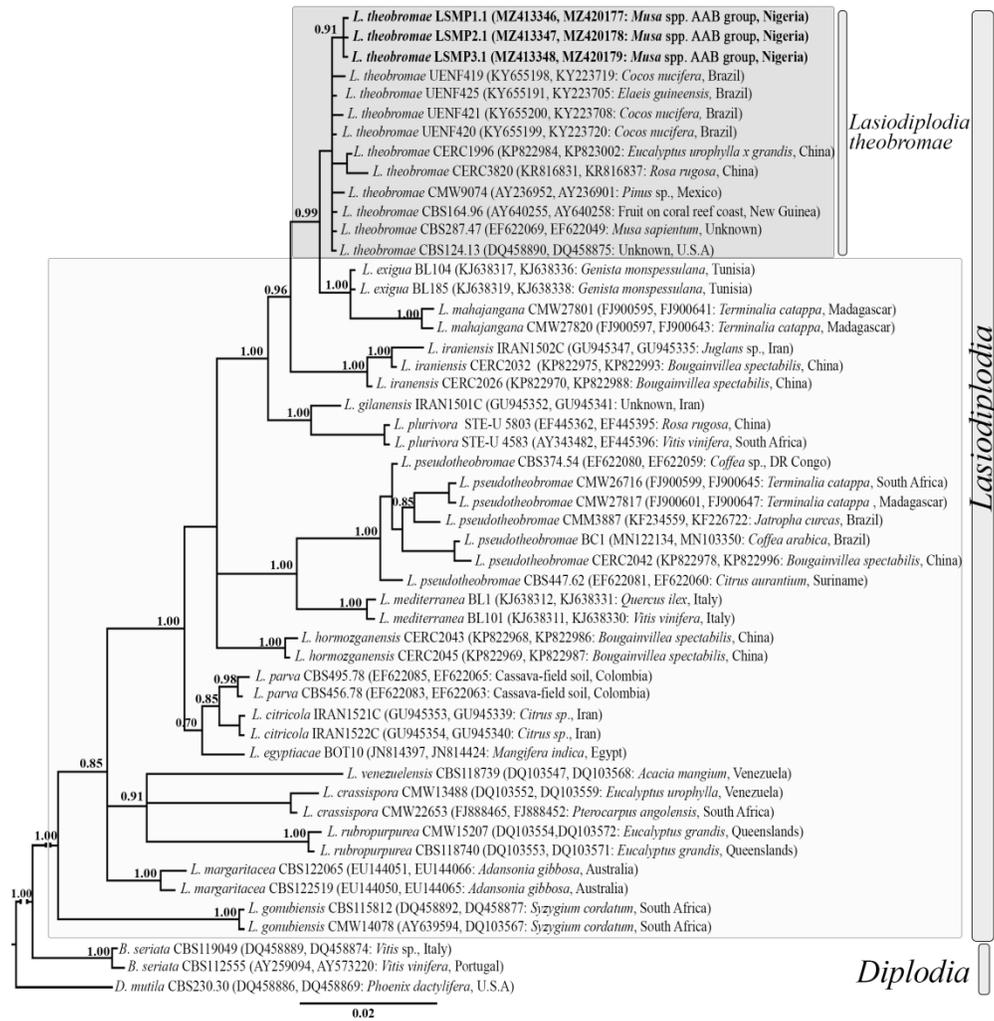
82

83 **Supplementary Figure S2.** Phylogenetic tree of *Lasiodiplodia* isolates from plantains (*Musa*
84 spp. AAB group) plants in Nigeria and related species based on the Bayesian Inference of a
85 combined internal transcribed spacer (ITS) and translation elongation factor 1-alpha (TEF 1-
86 α). The posterior probability is indicated near the branch nodes. *Diplodia mutila* CBS 230.30
87 is used as outgroup. Isolates marked in bold represent those obtained from plantains (*Musa*
88 spp. AAB group).



Supplementary Figure S1. Plants with fingers and peduncles rot, dieback symptoms caused by *Lasiodiplodia theobromae* (A-C), Pseudostem cross-section of infected plant (D), cultures of isolates on PDA (E-F), spores' morphology (G), and the rot caused by *L. theobromae* following the pathogenicity test on fruits (H) and pseudostems of plants (I). Scale bars: D-F, H-I = 2 cm; G = 10 μ m

209x124mm (300 x 300 DPI)



Supplementary Figure S2. Phylogenetic tree of *Lasiodiplodia* isolates from plantains (*Musa* spp. AAB group) plants in Nigeria and related species based on the Bayesian Inference of a combined internal transcribed spacer (ITS) and translation elongation factor 1-alpha (TEF 1- α). The posterior probability is indicated near the branch nodes. *Diplodia mutila* CBS 230.30 is used as outgroup. Isolates marked in bold represent those obtained from plantains (*Musa* spp. AAB group).

209x215mm (300 x 300 DPI)